

The Potential Effect of Ferric Iron and Bacteria Levels on Mold Density in Soil



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Abstract

Fungi are essential organisms in soil because they decompose dead matter and replenish the soil with nutrients. Mold, a form of fungi, utilizes ferric iron (Fe^{+3}) in a process that creates energy which allows it to grow and reproduce. Therefore, when ferric iron levels drop in the soil mold levels also decrease. One thing that can cause this to happen is when bacteria convert ferric iron (Fe^{+3}) to ferrous iron (Fe^{+2}), a form of iron mold cannot use. When the 2014 E.S.S.R.E. Biota Survey revealed varied and unexpected mold densities across the microclimate research sites, we hypothesized it was due to high levels of bacteria in the soil. Soil core samples were extracted from quadrants 2 and 4 of Site 4 and tested for ferric iron levels while serial dilutions were also performed to determine the densities of mold and bacteria in those soil samples. Although some of the results showed the expected correlations between the amount of bacteria, mold, and iron in the soil, these correlations had low statistical significance. To extend this study, we would explore potential correlations between light, water, bacteria densities, and mold densities in Site 4 due to the fact that light and water also vary in this location significantly and are critical environmental factors that influence the viability of these two critical soil microbes.

Introduction

Molds play an important role in soil health by decomposing and preventing the buildup of organic matter. They thrive in moist, warm soil (North Carolina Department of Health and Human Services, 2014), and when there is a lower density of mold in the soil, matter is not decomposed efficiently, causing plants dependent on these released nutrients to suffer. One factor that influences the health of soil molds is the amount of Fe^{+3} present. Mold utilizes ferric iron (Fe^{+3}) in its mitochondria to aid in growth and reproduction (Isaac, 1997); therefore, when there is a lack of ferric iron in the soil, mold cannot grow and by extension reproduce, depleting the levels of mold in that particular site.

One of the environmental factors that can influence Fe^{+3} levels in the soil is the amount of bacteria living there, specifically members of the genera *Pseudomonas*, *Bacillus*, *Bacteroides*, and *Desulfovibrio*. Species of these bacteria groups can reduce Fe^{+3} to Fe^{+2} , thereby making the Fe^{+3} which molds need less available. Ferric iron reduction is most commonly seen in the neutrophilic, heterotrophic bacteria of these groups (Johnson and McGinness, 1991), and while the causes of ferric iron reduction are not fully known, there is evidence that bacteria may induce ferric iron reduction by using ferric iron as a hydrogen sink to produce energy (Johnson and McGinness, 1991). Regardless, where there are high levels of these bacteria in the soil, there can be low levels of ferric iron and, therefore, less mold present.

Based on the findings of the E.S.S.R.E 2014 Biota survey (E.S.S.R.E, 2014), we observed that mold density in the soil of microclimate Site 4 (4,808.33 /cc) was significantly lower than those in the other three microclimate sites. But given that Site 4 is a partial wetlands, this finding is highly unexpected. Upon examining the rest of the survey data, though, we noticed that the Fe^{+3} levels in Site 4 were also significantly lower (11.04 ppm) while the bacteria density was unusually high (12,174,166.67 /cc). Therefore, in this experiment we investigated the correlations between the quantities of bacteria, mold, and Fe^{+3} present in different parts of Site 4. We hypothesized that the higher amounts of bacteria in Site 4 are inversely affecting the amount of ferric iron and mold in the soil because of bacteria's ability to convert ferric iron into ferrous iron.

Methods

For 3 consecutive days 5 separate soil samples each 15 cm deep with a diameter of 2 cm were randomly extracted from both quadrant 2 and quadrant 4 In E.S.S.R.E. Site 4 (E.S.S.R.E., 2001) for a total of 10 samples per day. All soil samples were tested for Fe^{+3} (ppm) using The LaMotte Model STH-14 Series Test Kit while simultaneously each sample was serially diluted to 10⁻⁴ and 100 μl aliquots of the 10⁻¹ and 10⁻² dilutions were plated on separate 3M Petrifilm™ Yeast and Mold Count agar sheets and 100 μl aliquots of the 10⁻², 10⁻³ and 10⁻⁴ dilutions were plated on separate 3M Petrifilm™ Aerobic Count Plates agar sheets. All plates

were incubated at room temperature for 72 hours, and then the density of mold (#/cm³) and bacteria (#/cm³) in each soil sample was calculated.

Results

Table #1

Data Averages

	Quadrant 2			Quadrant 4		
	Iron (ppm)	Mold (#/cm ³)	Bacteria (#/cm ³)	Iron (ppm)	Mold (#/cm ³)	Bacteria (#/cm ³)
Day 1	8.25	2960	17262000	11	18400	26300000
Day 2	15.5	2100	568000	7.25	4220	1754000
Day 3	6.25	2875	2746000	13.5	5400	1550000

Table 1 indicates the correlations observed between iron levels and mold and bacteria densities in the soils sampled over the course of the 3 days of the experiment.

Table #2

p-values

	Quad 2		Quad 4	
	Day 1- Day 2	Day 2- Day 3	Day 1- Day 2	Day 2- Day 3
Iron	0.576	0.576	0.2898	0.565
Mold	0.698	0.9237	0.203	0.995
Bacteria	0.3346	0.4373	0.0296	0.821

Table 2 values determined by t-testing

Figure #1

The effect of the amount of Iron on the amount of Bacteria.

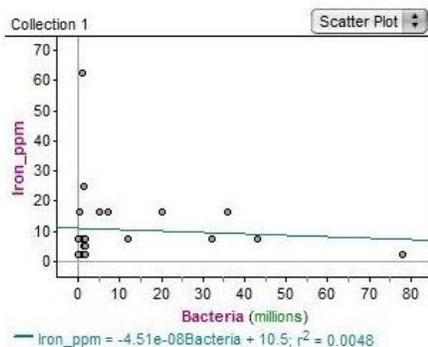


Figure 2 shows that the relationship between the amount of bacteria and amount of iron as a negative correlation, meaning that as the bacteria levels go up the Ferric iron levels go down. The r² value of .0048 shows a very weak negative correlation between the levels of bacteria and iron.

Figure #2

The effect of the amount of Iron on the amount of Mold.

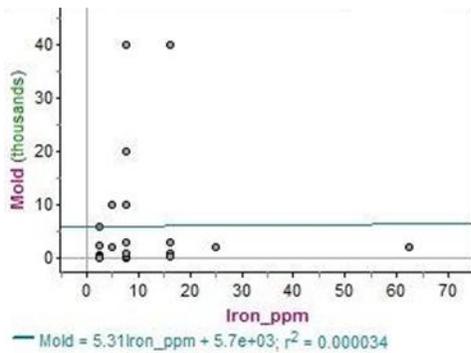


Figure 3 shows a positive correlation between the amount of mold and iron, meaning that as the iron levels go up the mold levels also go up. The r^2 value of .000034 shows a positive weak relationship between the levels of mold and iron.

Discussion

As Table 1 shows, the changes we observed in mold and bacteria densities and iron levels in the tested soil samples produced only partial support for our original hypothesis. While the changes in the bacteria densities show the expected inverse relationship with Fe^{+3} levels from one day to the next in quadrant 2, there is no statistical significance to this trend ($p = 0.576$ for iron and $p = 0.3346$ for bacteria). Furthermore, while the mold densities and Fe^{+3} levels in quadrant 4 do show the hypothesized direct correlation, they do so significantly only between Day 1 and Day 2 of the experiment ($p = 0.2898$ for iron and $p = 0.203$ for mold). Finally, as Figures 1 and 2 indicate, the trends we predicted were observed during our research, but the anticipated correlations are very weak at best ($r^2 = 0.0048$ for the relationship between bacteria and iron and $r^2 = 0.000034$ for the relationship between iron and mold). Hence, while there is data to support our original hypothesis, it is minimal at best.

However, the data in Table 1 do show two interesting trends. The first trend observed was that in quadrant 2, the bacteria densities fluctuated unsteadily while the mold densities remained at a steady level of about 2,500 per cm^3 over the course of three days. Conversely, in quadrant 4, it was the bacteria densities that stabilized at a steady level of 1.8 million per cm^3 while the mold densities fluctuated unsteadily. These two trends suggest that there is an alternative factor other than Fe^{+3} levels that may be at work in these two different quadrants.

One possibility is the physical environmental differences between quadrant 2 and 4 in Site 4. For example quadrant 4 lacks trees and is therefore exposed to more sunlight than in quadrant 2. In addition, quadrant 2 has a stream going through it, whereas quadrant 4 does not have any water flow in it. Given that both of these factors can play a role in the health of both bacteria (E.S.S.R.E., 2005) and molds (Nyberg, 1987), it would be interesting to explore the potential correlations between light, water, bacteria densities, and mold densities in Site 4. In the future, we would include gathering data on the levels of sunlight and moisture in both quadrants as well as measuring changes in the growth rates of mold and bacteria in these locations.

References

- General Descriptions of the ESSRE Survey Sites. (n.d.). Retrieved from Environmental Science Summer Research Experience website: <http://essre.rpcs.org/>
- Hochmuth, G. (n.d.). Iron (Fe) Nutrition of Plants. Retrieved from University of Florida website: <http://edis.ifas.ufl.edu>
- Isaac, S. (1997). Iron is relatively insoluble and often unavailable in the natural environment: how do fungi obtain sufficient supplies? In *Mycology Answers* (Vol. 11). Retrieved from <http://www.fungi4schools.org> (Reprinted from *Mycology Answers*, n.d.)
- Johnson, D. B., & McGinness, S. (n.d.). Ferric Iron Reduction by Acidophilic Heterotrophic Bacteria. *Applied and Environmental Microbiology*. Retrieved from <http://www.ncbi.nlm.nih.gov>
- North Carolina Department of Health and Human Services. (2014, June 20). Mold: Where It Can Grow. Retrieved July 22, 2014, from <http://epi.publichealth.nc.gov/oe/mold/grow.html>
- Nyberg, S. (1987). The Invasion of the Giant Spore. *SOLINET Preservation Program*. Retrieved from <http://www-sul.stanford.edu/tools/tutorials/html2.0/spore.html>
- Pongas, G. N., Ben-Ami, R., Lewis, R. E., Walsh, T. J., & Kontoyiannis, D. P. (2009). Culture Medium Composition Affects the Lethality of *Cunninghamella bertholletiae* in a Fly Model of Mucormycosis. *American Society for Microbiology*. Retrieved from <http://www.ncbi.nlm.nih.gov>
- Shulte, E.E. (2014). Soil and Applied Iron. *University of Wisconsin Extension*. Retrieved from <http://corn.agronomy.wisc.edu>

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